Generation, Visualisation and Analysis of Multiple Sequence Alignments

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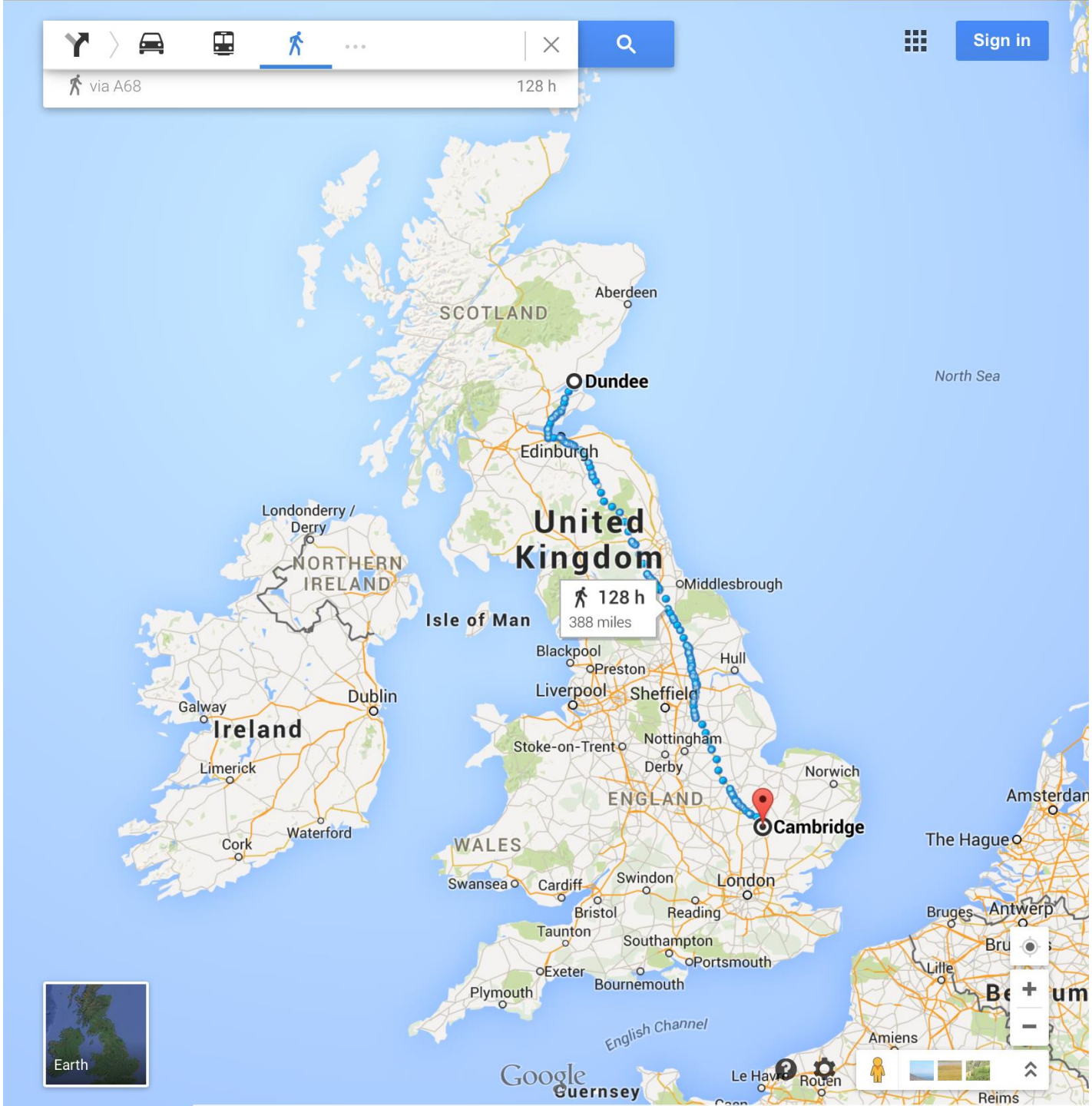
Jalview Training Course – May 2015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Leven
Elie
Crail
Anstruther
Buckhaven
Thornton
Ballingry
Cardenden
Kelty
Lochgelly
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Mitchelston Industrial Estate
Kirkcaldy
Dollar
Alva
Tillicoultry
Tullibody
Alloa
Oakley
Townhill
Dunfermline
Kincardine
Kinghorn
Burntisland
North Berwick
Gullane
Rosyth
Dalgety Bay
Carronshore
Denny
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Bo'ness
Bonnybridge
Falkirk
Polmont
Queensferry
Shieldhill
Linlithgow
Kirkliston
Leith
Longniddry
East Linton
Haddington
Tranent
Musselburgh
Corstorphine
Newington
Slamannan
Broxburn
Ratho
Ormiston
Dalkeith
Bathgate
Livingston
Currie
Balerno
Straiton
Caldercruix
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M90
A9
A85
A91
A92
A915
A902
A720
A71
M9
M876
A977
A876
A985
M8

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Top University for Biological Sciences in the UK REF 2014
Highest ISI citation rate of Life Sciences Departments in Europe
22 Principal Investigators in top 1% of cited scientists
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School of Life Sciences Research

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One floor: Research Division of Computational Biology (currently 6 groups)

Recruiting 5 new research groups in Computational Biology over next 5 years

What is multiple sequence alignment (MSA)?

- Alignment of three or more sequences
- What does one look like?

Example Multiple Sequence alignment of 27 SH2 domains

Why are Protein (multiple) Sequence Alignments Useful?

- Link proteins at the amino acid level
- Allow identification of conserved features
- Allow prediction of functionally important residues
- Basis for phylogenetic tree construction
- Basis for sensitive profile-based sequence database searching
- Basis for training many methods to predict features from sequence – e.g. secondary structure

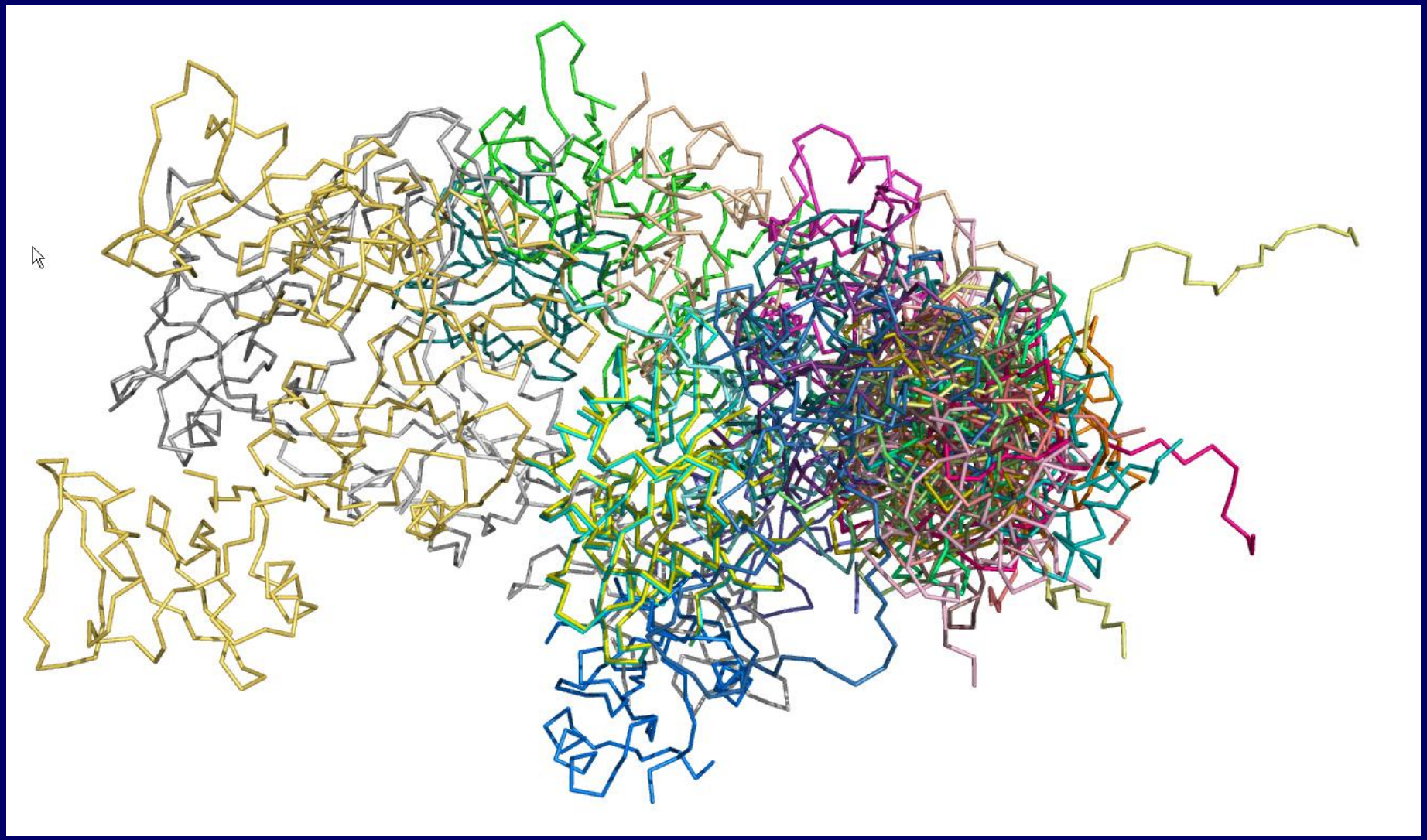
- Standard way of describing and illustrating features of protein sequences and their relationships in publications

Link proteins at the amino acid level

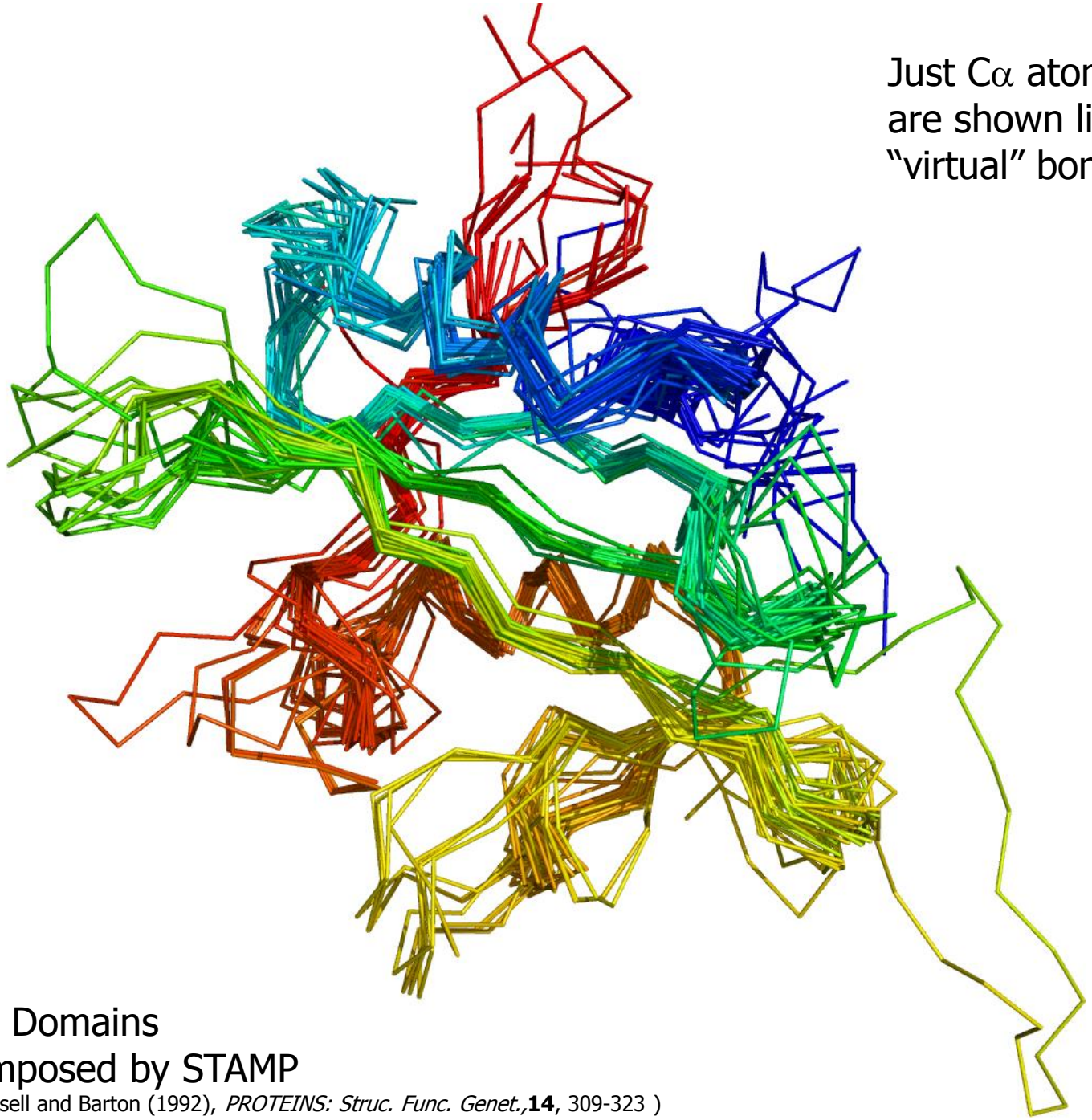
What does this mean?

Example sequence alignment of SH2 domains

From the *three-dimensional* structures of the proteins



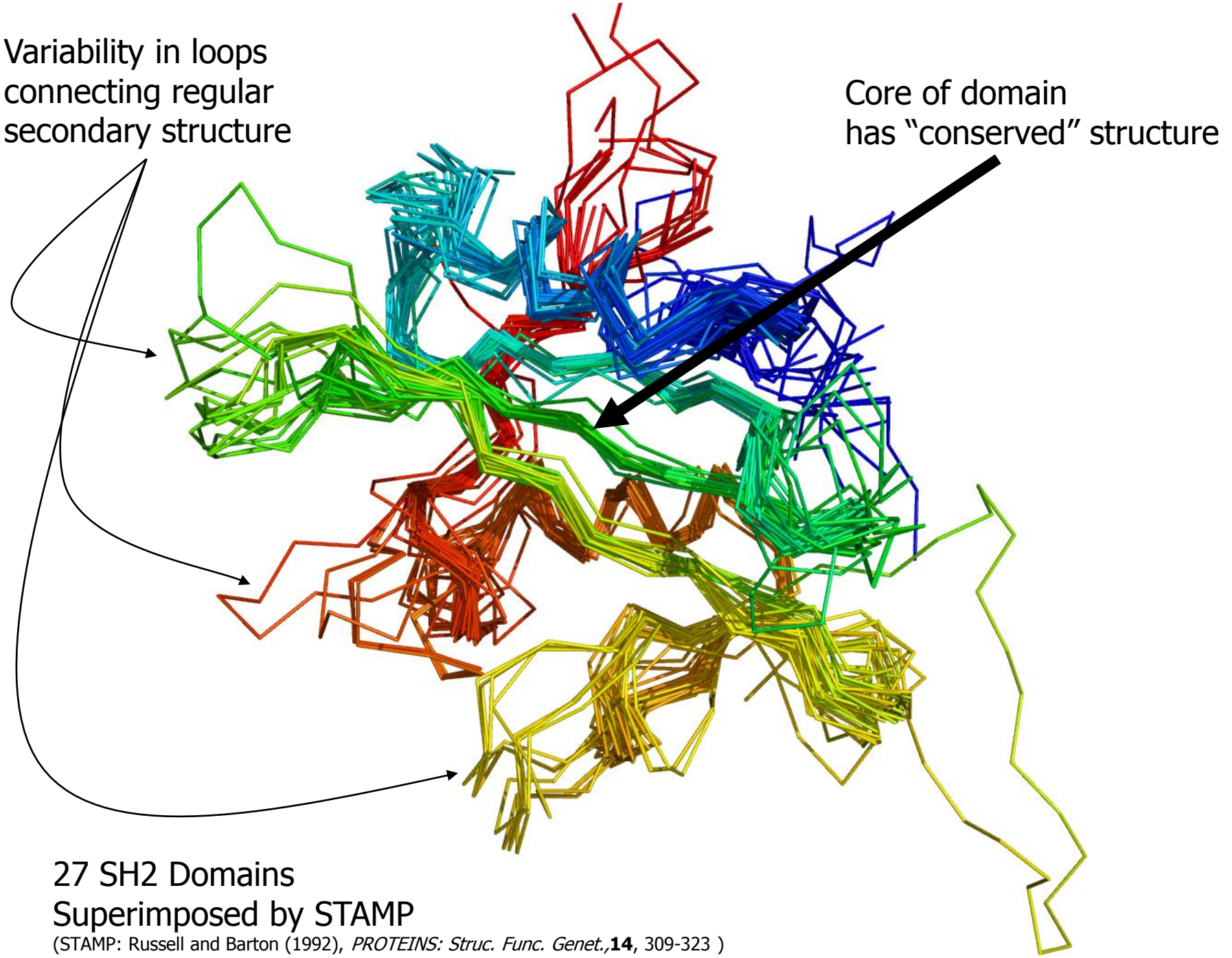
22 SH2 domain structures as they are if just loaded into PyMol from the PDB



Just Cα atoms
are shown linked by
"virtual" bonds

27 SH2 Domains
Superimposed by STAMP

(STAMP: Russell and Barton (1992), *PROTEINS: Struc. Func. Genet.,***14**, 309-323)



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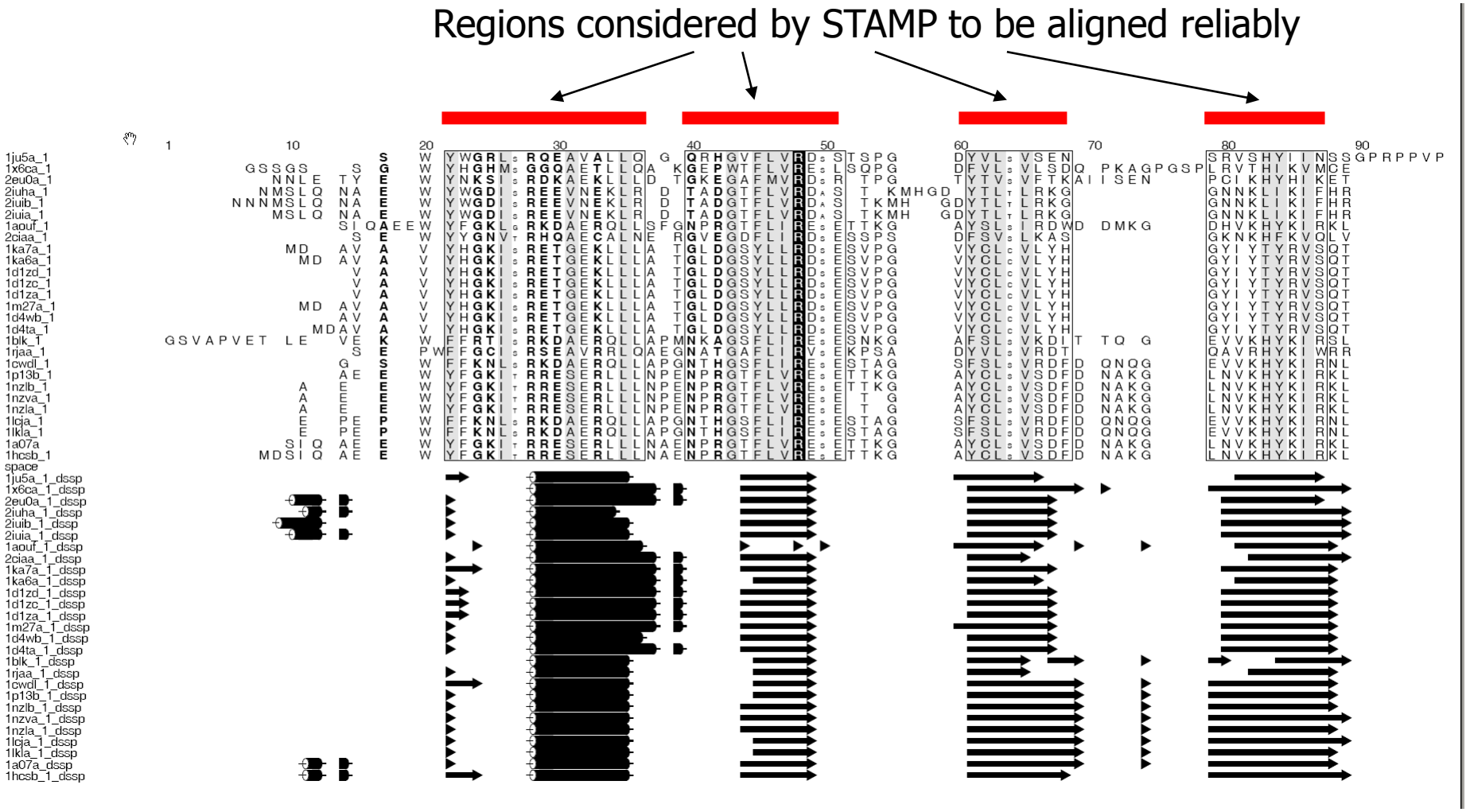
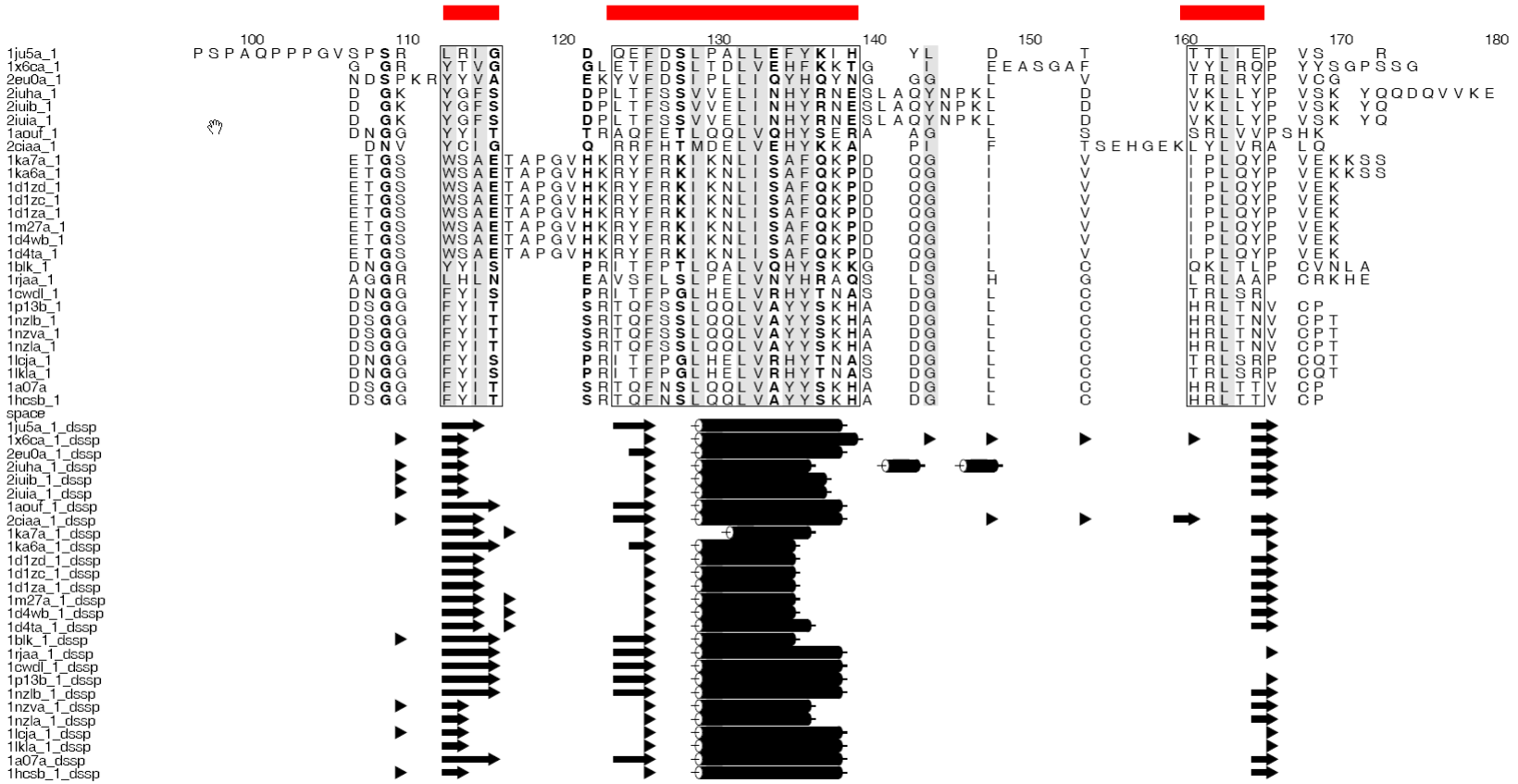


Structural alignment of 27 SH2 domains showing secondary structure – Part I

STAMP alignment – Alscript display



Structural alignment of 27 SH2 domains showing secondary structure – Part II

STAMP alignment – Alscript display

How are MSAs generated when we just have sequences and no knowledge of 3D structure?

Multiple Sequence Alignment from the 1960/70s

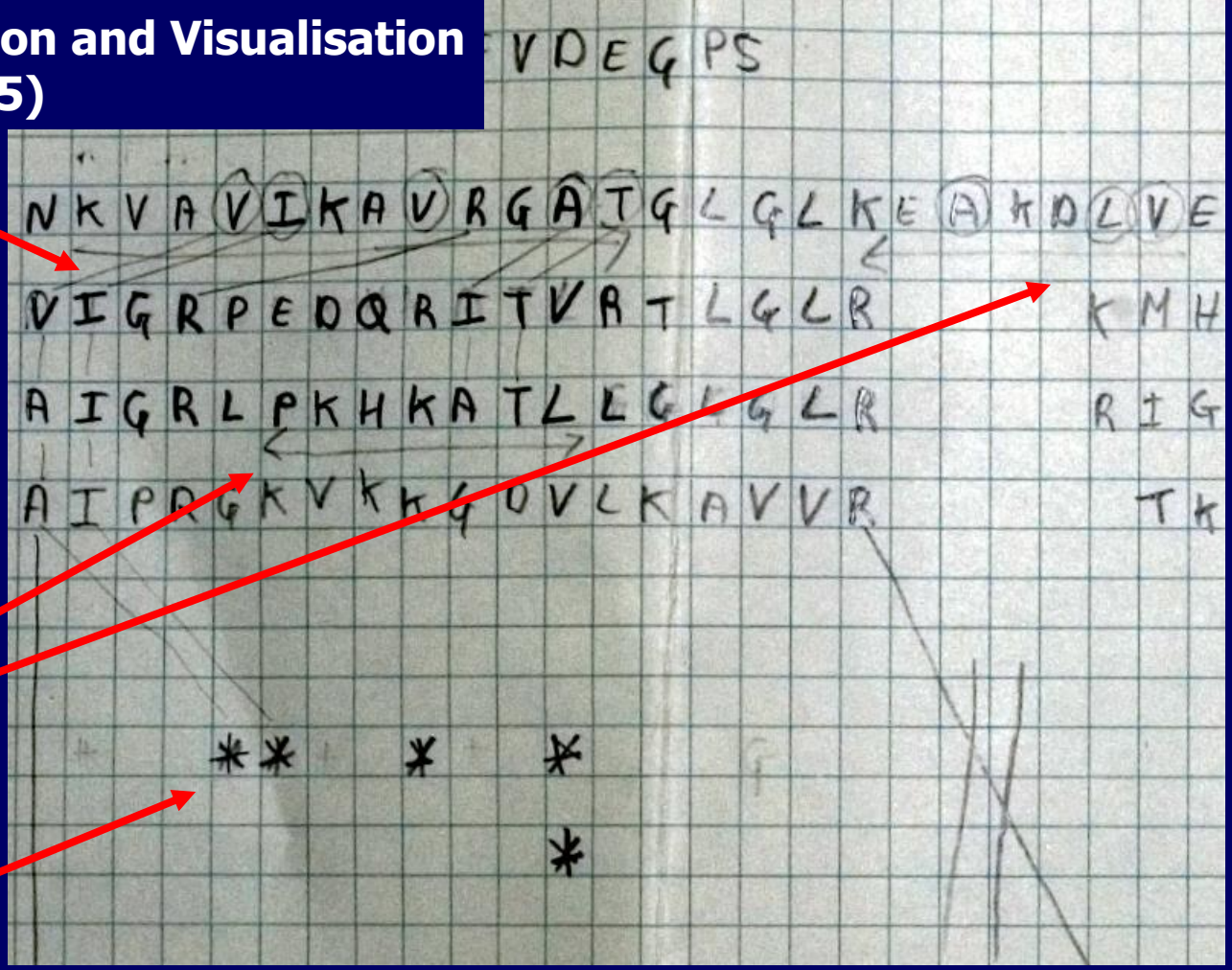
Courtesy of the University of Edinburgh

1984

- Some sequence alignment programs existed but common method was to…
- Align two sequences by writing the amino acid codes on squared paper then sliding them relative to each other to find a good match.
- Use scissors to deal with insertions/deletions
- Yes really!

Multiple Alignment Creation and Visualisation (1984/5)

"Trace"
(Alternative Alignment)



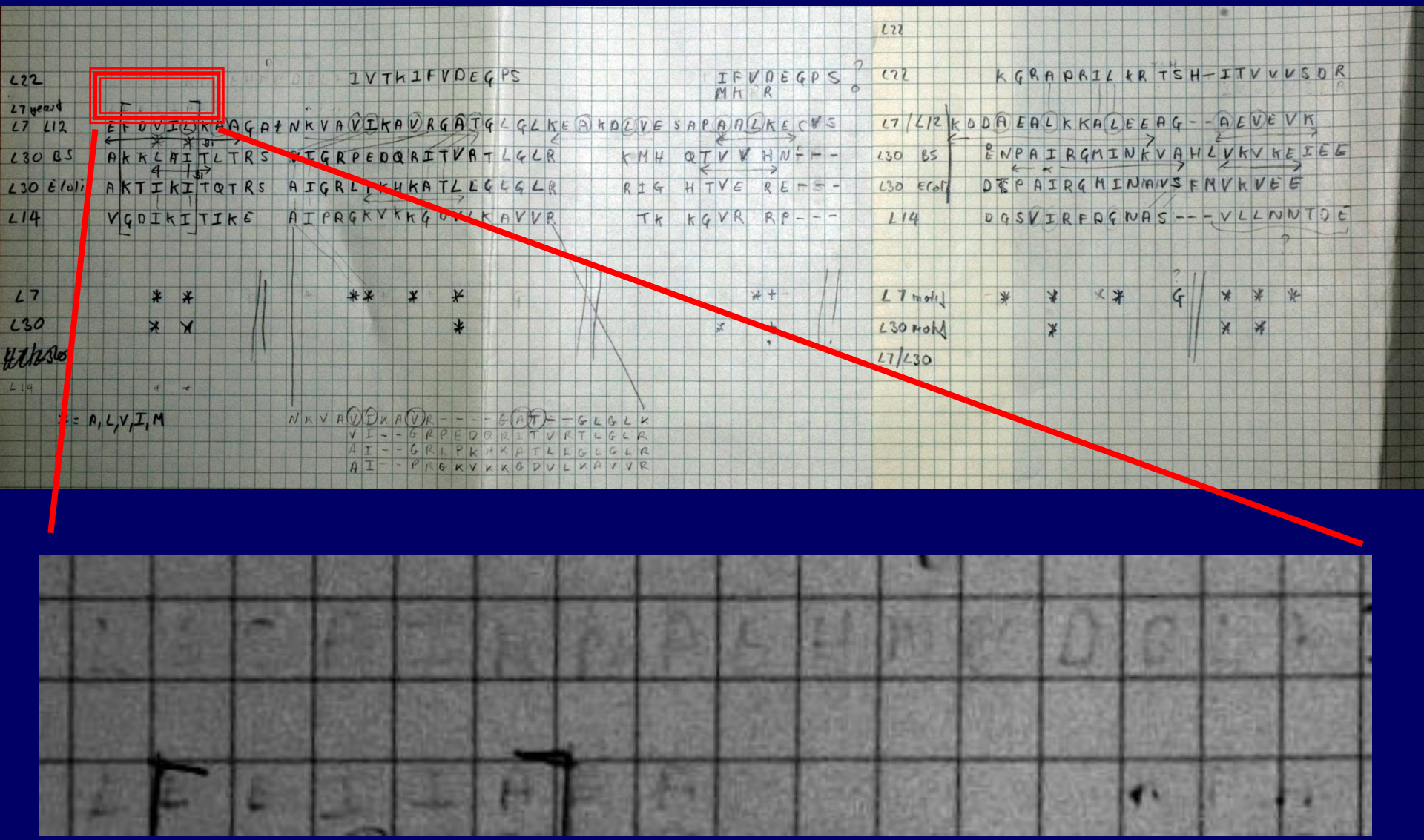
Location of Beta-Strand
(double headed arrows)

"*" ??

Re-drawn
Alternative Alignment

$* = A, L, V, I, M$

Multiple Sequence Alignment and Visualisation (1984/5)



How are MSAs generated?

Pair-wise sequence alignment

Alignment of *two* Protein Sequences -How?

- Need scoring scheme for matching amino acid residues.
- Need to cope with insertions and deletions (gaps or indels).
- Need algorithm to find 'best' alignment.
- Need some way of judging if the alignment is likely to be correct.

Protein Scoring Schemes

- A table of scores for aligning each possible amino acid pair.
- **Simplest scheme**, just scores 1 for identity and 0 for non identity.

- Better schemes weight similarities in amino acid properties or observed substitutions. For example, BLOSUM and PAM series. Virtually all of today's programs use these.

BLOSUM62 Matrix

	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V	B	Z	X	*
A	4	-1	-2	-2	0	-1	-1	0	-2	-1	-1	-1	-1	-2	-1	1	0	-3	-2	0	-2	-1	0	-4
R	-1	5	0	-2	-3	1	0	-2	0	-3	-2	2	-1	-3	-2	-1	-1	-3	-2	-3	-1	0	-1	-4
N	-2	0	6	1	-3	0	0	0	1	-3	-3	0	-2	-3	-2	1	0	-4	-2	-3	3	0	-1	-4
D	-2	-2	1	6	-3	0	2	-1	-1	-3	-4	-1	-3	-3	-1	0	-1	-4	-3	-3	4	1	-1	-4
C	0	-3	-3	-3	9	-3	-4	-3	-3	-1	-1	-3	-1	-2	-3	-1	-1	-2	-2	-1	-3	-3	-2	-4
Q	-1	1	0	0	-3	5	2	-2	0	-3	-2	1	0	-3	-1	0	-1	-2	-1	-2	0	3	-1	-4
E	-1	0	0	2	-4	2	5	-2	0	-3	-3	1	-2	-3	-1	0	-1	-3	-2	-2	1	4	-1	-4
G	0	-2	0	-1	-3	-2	-2	6	-2	-4	-4	-2	-3	-3	-2	0	-2	-2	-3	-3	-1	-2	-1	-4
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L	-1	-2	-3	-4	-1	-2	-3	-4	-3	2	4	-2	2	0	-3	-2	-1	-2	-1	1	-4	-3	-1	-4
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M	-1	-1	-2	-3	-1	0	-2	-3	-2	1	2	-1	5	0	-2	-1	-1	-1	-1	1	-3	-1	-1	-4
F	-2	-3	-3	-3	-2	-3	-3	-3	-1	0	0	-3	0	6	-4	-2	-2	1	3	-1	-3	-3	-1	-4
P	-1	-2	-2	-1	-3	-1	-1	-2	-2	-3	-3	-1	-2	-4	7	-1	-1	-4	-3	-2	-2	-1	-2	-4
S	1	-1	1	0	-1	0	0	0	-1	-2	-2	0	-1	-2	-1	4	1	-3	-2	-2	0	0	0	-4
T	0	-1	0	-1	-1	-1	-1	-2	-2	-1	-1	-1	-1	-2	-1	1	5	-2	-2	0	-1	-1	0	-4
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V	0	-3	-3	-3	-1	-2	-2	-3	-3	3	1	-2	1	-1	-2	-2	0	-3	-1	4	-3	-2	-1	-4
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Z	-1	0	0	1	-3	3	4	-2	0	-3	-3	1	-1	-3	-1	0	-1	-3	-2	-2	1	4	-1	-4
X	0	-1	-1	-1	-2	-1	-1	-1	-1	-1	-1	-1	-1	-1	-2	0	0	-2	-1	-1	-1	-1	-1	-4
*	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	-4	1

BLOSUM62 is a log-score matrix – more on this later...

Gap Penalties

- Score for aligning a residue or residues in one protein to a gap in the other.
- Most usual form:
  penalty = ul + v
- where l is the length of the gap and u and v are constants.
- u is often called the gap extension penalty, v, the gap creation penalty.

Finding the 'best' alignment

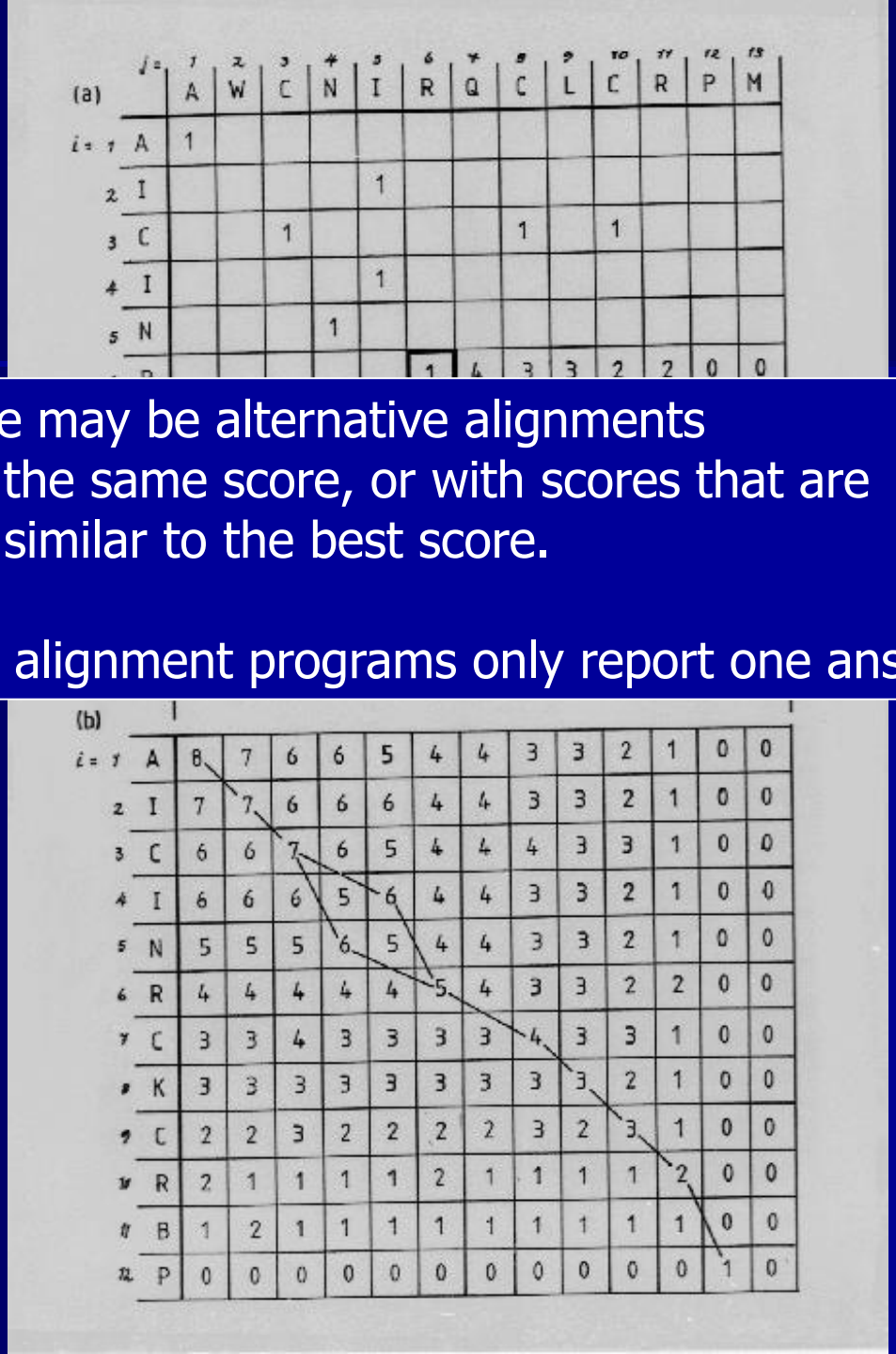
- The mathematically best alignment is the one that gives the highest score when the amino acids of the two proteins are aligned, taking account of any gaps.
- *This alignment is not necessarily the one that is biologically meaningful.* (more on this later)

Finding the best alignment

- Naïve way would be to generate all possible alignments of the two sequences, then take the one with the highest score according to the BLOSUM matrix.
- But… for two sequences of 100 amino acids, there are > 10^{75} possible alignments…

Dynamic Programming

- Trick to avoid having to generate all possible alignments.
- First introduced in molecular biology by Needleman and Wunsch (1970).
- Many variations on the theme.
- Basis of (nearly) all sequence alignment programs.
- Finds the mathematically 'best' score for alignment of two sequences of length *M* and *N* in *MN* steps.



(a)

		j =	1 A	2 W	3 C	4 N	5 I	6 R	7 Q	8 C	9 L	10 C	11 R	12 P	13 M
i = 1	A		1												
2	I						1								
3	C				1					1		1			
4	I						1								
5	N					1									
								1	4	3	3	2	2	0	0

There may be alternative alignments with the same score, or with scores that are very similar to the best score.

Most alignment programs only report one answer…

(b)

i = 1	A	8	7	6	6	5	4	4	3	3	2	1	0	0
2	I	7	7	6	6	6	4	4	3	3	2	1	0	0
3	C	6	6	7	6	5	4	4	4	3	3	1	0	0
4	I	6	6	6	5	6	4	4	3	3	2	1	0	0
5	N	5	5	5	6	5	4	4	3	3	2	1	0	0
6	R	4	4	4	4	4	5	4	3	3	2	2	0	0
7	C	3	3	4	3	3	3	3	4	3	3	1	0	0
8	K	3	3	3	3	3	3	3	3	3	2	1	0	0
9	C	2	2	3	2	2	2	2	3	2	3	1	0	0
10	R	2	1	1	1	1	2	1	1	1	1	2	0	0
11	B	1	2	1	1	1	1	1	1	1	1	1	0	0
12	P	0	0	0	0	0	0	0	0	0	0	0	1	0

From:
Needleman &
Wunsch (1970)

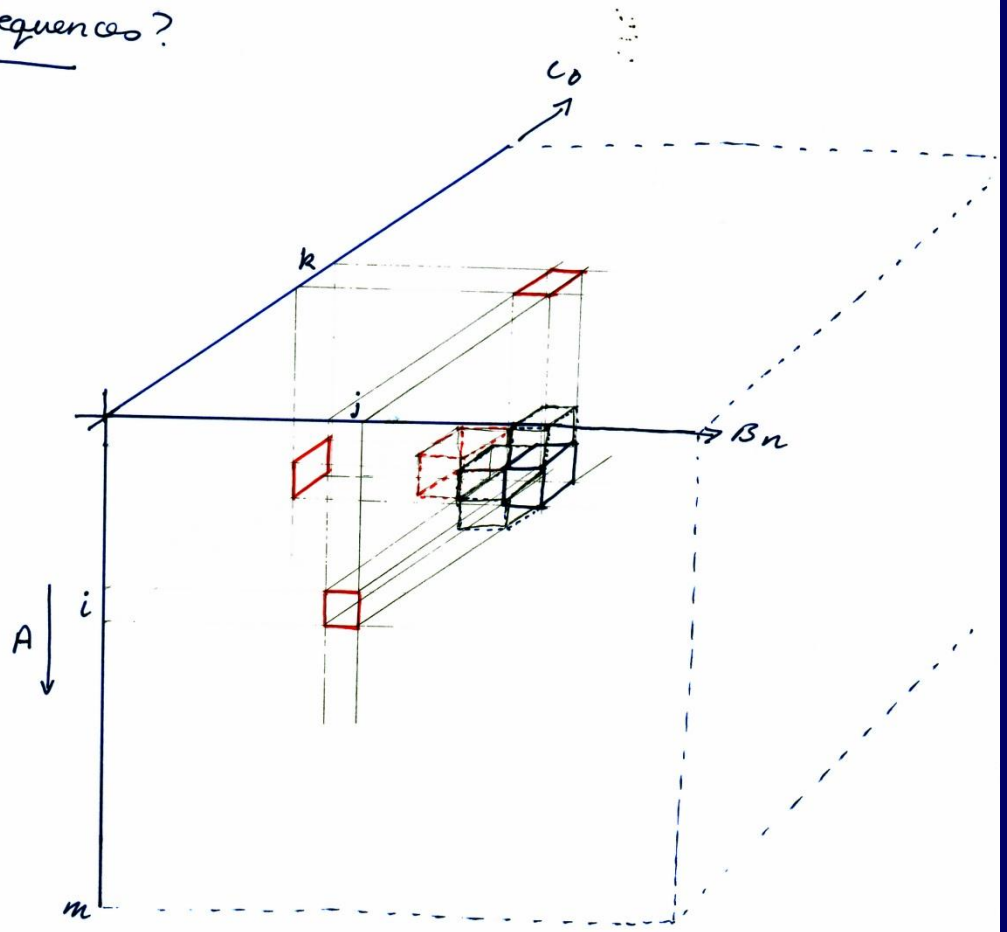
Multiple Sequence Alignment

Extension of two-sequence dynamic programming

For three sequences

Need a 3-dimensional array

3- Sequences?



3- Way Dynamic programming.

- gets complicated! - AND HARD TO DRAW!

BUT NOT IMPOSSIBLE

For *n* sequences?

Need an *n*-dimensional array…

Dynamic programming for >3 sequences

- Need an N-dimensional "hypercube"
- Very complex
- Very memory intensive
- Very CPU intensive

- e.g. to align 100 sequences of length 100.
  Need to store 100^{100} bytes.
  i.e. A BIG NUMBER!

- NOT PRACTICAL

Alternatives to dynamic programming

- Genetic Algorithms
  - Simulate process of "evolution", but for protein sequence alignments
  - Mutation/recombination of alignments
  - Has been implemented in the SAGA program
- STILL IMPRACTICAL for most use.

Hierarchical multiple alignment

- Compare all pairs of sequences
- Generate a guide tree or dendrogram
- Follow tree from leaves to root, building the alignment as you go.

- Virtually all current programs use this approach

- Most popular program is CLUSTAL. More recent and often more accurate programs are:
  - probcons, mafft and muscle…

Example: Alignment of 7 sequences with identifier codes HAHU, HBHU etc.

"Single linkage" dendrogram.

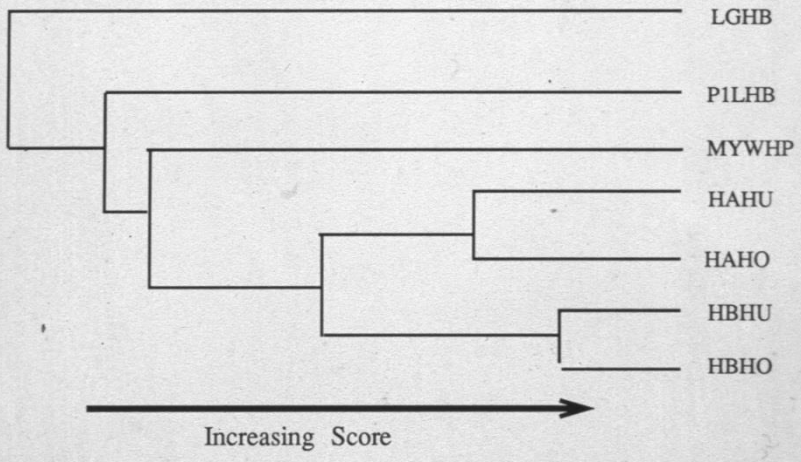
1. Most similar pair put together
2. Next most similar pair
3. and so on…

When one or both halves of a pair is an existing alignment, then do *profile* comparison.

PAIRWISE SCORES

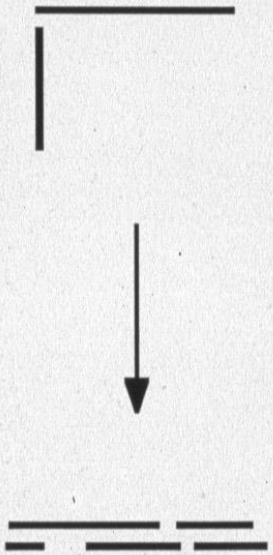
	HAHU	HBHU	HAHO	HBHO	MYWHP	P1LHB	LGHB
HAHU							
HBHU	21.1						
HAHO	32.9	19.7					
HBHO	20.7	39.0	20.4				
MYWHP	11.0	9.8	10.3	9.7			
P1LHB	9.3	8.6	9.6	8.4	7.0		
LGHB	7.1	7.3	7.5	7.4	7.3	4.3	

CLUSTER ANALYSIS



DENDROGRAM

Sequence vs Sequence

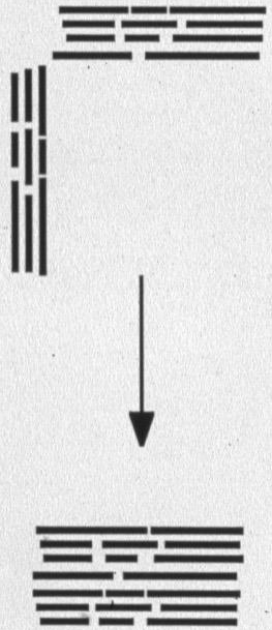


Pairwise Alignment

Alignment vs Sequence

Profile Alignment

Alignment vs Alignment



Profile-Profile Alignment

What is a Profile?

Making a profile: Given a multiple sequence alignment…

Multiple Alignment Frequency Profile

Amino Acid Types

Gaps

Position in Alignment

728 Glycines

13 Gaps

48 Prolines

11th position in alignment

Posn:	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V	B	Z	X	-
1:	95	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
2:	4	0	1	1	0	0	0	1	0	1	0	2	0	0	755	3	2	0	0	0	0	0	0	0
3:	161	19	15	120	6	39	142	4	4	6	28	113	3	3	38	60	48	1	0	19	0	0	0	0
4:	81	49	27	31	6	29	58	5	16	3	0	21	4	24	1	393	91	2	19	8	0	0	0	3
5:	4	5	0	0	12	0	6	0	0	176	60	19	30	162	0	1	16	66	26	353	0	0	0	0
6:	8	0	38	891	0	10	5	1	7	0	0	7	0	0	0	1	1	0	1	1	0	0	0	0
7:	9	[illegible]	[illegible]	[illegible]	[illegible]	[illegible]	0	0	0	4	38	0	0	9	0	1	0	956	27	[illegible]	[illegible]	[illegible]	0	1
8:	6	[illegible]	[illegible]	[illegible]	[illegible]	[illegible]	1	4	2	4	0	17	0	0	84	22	24	1	1	[illegible]	[illegible]	[illegible]	0	5
9:	45	17	106	145	4	79	196	16	13	5	24	175	7	3	8	48	115	1	4	34	0	0	0	5
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[illegible]	409	9	9	0	56	5	0	[illegible]	7	45	35	52	9	15	[illegible]	71	72	5	116	98	0	0	0	[illegible]
13:	13	1	3	0	5	4	0	1	0	178	70	2	12	3	0	1	5	0	0	770	0	0	0	2
14:	38	18	52	5	2	8	1	66	17	6	16	30	0	1	15	141	625	4	4	21	0	0	0	0
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20:	51	17	17	4	1	23	20	846	23	3	11	48	1	1	3	66	2	0	1	2	0	0	0	1
21:	58	46	118	50	3	235	46	48	50	13	32	53	39	6	25	215	40	7	33	22	0	0	1	4
22:	3	0	4	1	1123	0	0	0	0	0	4	0	1	0	1	6	2	1	0	1	0	0	0	3
23:	51	7	32	5	1	2	5	996	1	2	3	8	1	0	3	26	1	1	3	0	0	0	0	3
24:	81	1	4	2	59	1	2	53	0	3	11	0	0	0	0	908	18	3	0	5	0	0	0	2
25:	6	1	0	7	1042	0	0	13	1	1	3	0	0	0	0	76	0	0	0	3	0	0	0	2

Convert frequency profile into log-odds profile

In words:

log (proportion of a particular amino acid type at a position
divided by
proportion of that amino acid in the whole alignment)

Results in a
Negative number when amino acid is less common
at a position than in the alignment as a whole.
or a
Positive number if the amino acid is more common at
a position than in the alignment as a whole.

Conversion is usually more complex than this because you have to deal with the absence of amino acids at a position. This is done by taking background scores from a pair-score matrix like BLOSUM.

Example

- Alignment of 30 sequences each of 100 amino acids to give a total of 3000 amino acids
- Position 97 of the alignment has 20 prolines
- There are 300 prolines in the alignment as a whole
- (20/30)/(300/3000) = 20/3 = 6.67
- log(6.67) = 0.82
- So, score in profile for proline at position 97 is 0.82

- This is sometimes called a log-likelihood ratio

Log score profile

- In this example log values are multiplied by 100 to allow for integer arithmetic which is faster on most computers.

Pos	AA	Freq:	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V	B	Z	X	-	^	:	!
1	A	86:	755	-281	-181	-281	-181	-181	-181	-81	-281	-181	-281	-181	-181	-381	-181	18	-81	-381	-281	-81	-281	-181	-181	200	0	1200	-3000
2	P	722:	-176	-113	-324	-324	-210	78	77	-454	-20	-367	-212	-291	-114	-217	1486	-161	-127	-220	-117	-113	-11	78	-16	200	0	1200	-3000
3	A	779:	266	-224	-141	40	-398	-20	271	-602	-180	-299	-212	137	-46	-453	-59	54	86	-721	-175	-276	-9	61	-83	200	0	1200	-3000
4	A	805:	219	-8													-469	475	194	-439	-70	-398	-16	19	-69	200	0	1200	-3000
5	V	849:	-346	-389													-275	-496	-107	115	329	316	-309	-215	-90	200	0	1200	-3000
6	D	870:	-183	-36													22	-133	-219	-331	-317	-619	558	224	32	200	0	1200	-3000
7	W	926:	-173	-481													-211	-485	-87	1734	446	-372	-292	-38	-89	200	0	1200	-3000
8	R	927:	-143	947	-256	-304	-608	-179	-156	-514	-88	-389	-130	231	-36	-139	68	17	-82	-86	-153	-341	51	137	47	200	0	1200	-3000
9	A	927:	-29	-246	265	271	-454	78	341	-201	-53	-553	-337	151	-120	-327	-141	85	42	-292	-276	-335	116	111	-71	200	0	1200	-3000
10	R	862:	-77	131	20	-263	44	19	199	-668	208	-447	-49	366	-139	-12	375	-9	-165	-50	113	-391	-19	55	-55	200	0	1200	-3000
11	G	816:	-19	-258	234	79	-331	-270	-66	650	45	-441	-379	-94	-70	-231	-103	141	-143	-26	-165	-662	89	11	-34	200	0	1200	-3000
12	A	816:	245	-296	-371	-235	-206	-460	-182	-325	-171	-43	6	-151	-48	67	-433	93	-5	-191	166	-13	-210	-159	-121	200	0	1200	-3000
13	V	734:	11	-508	-342	-290	-315	-529	-217	-726	-270	595	372	-479	360	99	-281	-431	-94	-214	-36	641	-277	-192	-32	200	0	1200	-3000
14	T	734:	-54	-185	70	-275	-508	-363	-126	-240	-44	-338	-165	-209	-96	-189	-107	311	565	-195	-37	-297	7	-45	-32	200	0	1200	-3000
15	A	622:	152	-139	-165	-182	-262	-154	197	-401	-107	-339	-347	-92	-98	-160	561	111	-19	-297	-51	-439	-77	18	-113	200	0	1200	-3000
16	V	471:	83	-304	-406	-344	-152	-260	-286	-678	-354	504	-4	-264	109	-137	-100	-253	107	-317	-139	607	-338	-263	-114	200	0	1200	-3000
17	K	416:	-206	449	-268	-231	-259	14	19	-589	-3	-255												85	-63	200	0	1200	-3000
18	D	323:	-298	-15	449	504	-496	-318	47	-472	141	-261												53	72	200	0	1200	-3000
19	Q	209:	-104	24	-120	-88	-293	698	102	-525	121	-148												272	64	200	0	1200	-3000
20	G	198:	18	-77	-17	-130	-534	-113	-178	364	-60	-185	-231	-13	-224	-210	-299	14	-181	-274	-188	-232	-65	-90	-130	200	0	1200	-3000
21	Q	106:	-130	30	-78	-21	-593	122	21	-513	-42	-183	-219	59	-113	-99	-32	21	50	-334	-102	-204	-77	-34	25	200	0	1200	-3000
22	C	35:	-47	-253	-52	-170	538	-150	-172	-201	-188	-115	-66	-180	-31	-168	-183	-36	-18	-245	-179	35	-148	162	-125	200	0	1200	-3000
23	G	21:	-77	-98	-51	-109	-172	-9	-66	-151	-51	-177	-161	-51	-72	-188	-140	-51	-61	-172	-82	-161	-93	-40	-114	200	0	1200	-3000
24	S	390:	-100	-341	53	-2	-82	90	25	-57	-153	-214	-333	-78	-160	-274	-163	380	110	-653	-204	-368	-69	-53	-117	200	0	1200	-3000
25	C	326:	-76	-215	-78	-167	1043	-137	-156	-230	-130	-116	-71	-140	-65	-121	-233	5	12	-336	-159	-137	-156	-144	-81	200	0	1200	-3000
26	W	326:	-258	-73	-133	-72	-238	37	-86	-147	-9	-78	-125	-85	50	-14	-180	-400	-228	1182	529	-183	-203	-33	221	200	0	1200	-3000

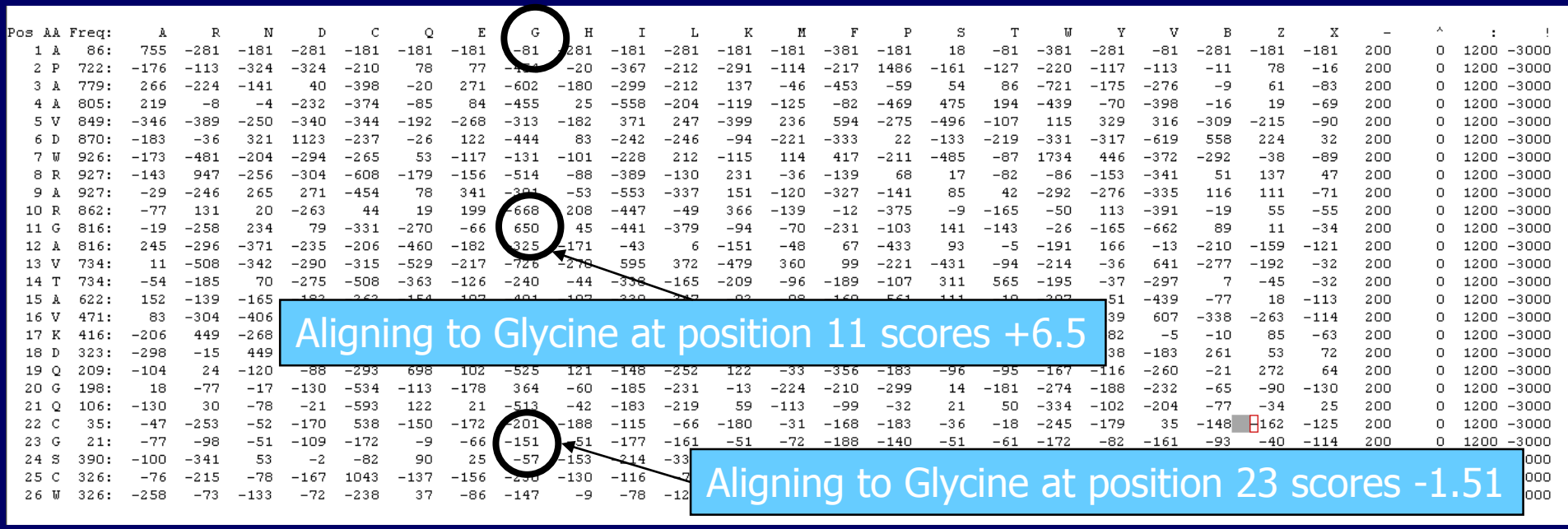
728 Glycines – gives score of +6.5

48 Prolines gives score of -1.03

How is a profile used in alignment?

- Rather than getting the score for aligning a particular residue at a position from the BLOSUM matrix take it from the profile.

Profiles give position-specific scoring



Pos	AA	Freq:	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V	B	Z	X	-	^	:	!
1	A	86:	755	-281	-181	-281	-181	-181	-181	-81	-281	-181	-281	-181	-181	-381	-181	18	-81	-381	-281	-81	-281	-181	-181	200	0	1200	-3000
2	P	722:	-176	-113	-324	-324	-210	78	77	-[illegible]	-20	-367	-212	-291	-114	-217	1486	-161	-127	-220	-117	-113	-11	78	-16	200	0	1200	-3000
3	A	779:	266	-224	-141	40	-398	-20	271	-602	-180	-299	-212	137	-46	-453	-59	54	86	-721	-175	-276	-9	61	-83	200	0	1200	-3000
4	A	805:	219	-8	-4	-232	-374	-85	84	-455	25	-558	-204	-119	-125	-82	-469	475	194	-439	-70	-398	-16	19	-69	200	0	1200	-3000
5	V	849:	-346	-389	-250	-340	-344	-192	-268	-313	-182	371	247	-399	236	594	-275	-496	-107	115	329	316	-309	-215	-90	200	0	1200	-3000
6	D	870:	-183	-36	321	1123	-237	-26	122	-444	83	-242	-246	-94	-221	-333	22	-133	-219	-331	-317	-619	558	224	32	200	0	1200	-3000
7	W	926:	-173	-481	-204	-294	-265	53	-117	-131	-101	-228	212	-115	114	417	-211	-485	-87	1734	446	-372	-292	-38	-89	200	0	1200	-3000
8	R	927:	-143	947	-256	-304	-608	-179	-156	-514	-88	-389	-130	231	-36	-139	68	17	-82	-86	-153	-341	51	137	47	200	0	1200	-3000
9	A	927:	-29	-246	265	271	-454	78	341	-[illegible]	-53	-553	-337	151	-120	-327	-141	85	42	-292	-276	-335	116	111	-71	200	0	1200	-3000
10	R	862:	-77	131	20	-263	44	19	199	-668	208	-447	-49	366	-139	-12	-375	-9	-165	-50	113	-391	-19	55	-55	200	0	1200	-3000
11	G	816:	-19	-258	234	79	-331	-270	-66	650	45	-441	-379	-94	-70	-231	-103	141	-143	-26	-165	-662	89	11	-34	200	0	1200	-3000
12	A	816:	245	-296	-371	-235	-206	-460	-182	325	-171	-43	6	-151	-48	67	-433	93	-5	-191	166	-13	-210	-159	-121	200	0	1200	-3000
13	V	734:	11	-508	-342	-290	-315	-529	-217	-726	-278	595	372	-479	360	99	-221	-431	-94	-214	-36	641	-277	-192	-32	200	0	1200	-3000
14	T	734:	-54	-185	70	-275	-508	-363	-126	-240	-44	-338	-165	-209	-96	-189	-107	311	565	-195	-37	-297	7	-45	-32	200	0	1200	-3000
15	A	622:	152	-139	-165	[illegible]																	51	-439	-77	18	-113	200	0	1200	-3000
16	V	471:	83	-304	-406	[illegible]																	39	607	-338	-263	-114	200	0	1200	-3000
17	K	416:	-206	449	-268	[illegible]																	82	-5	-10	85	-63	200	0	1200	-3000
18	D	323:	-298	-15	449	[illegible]																	38	-183	261	53	72	200	0	1200	-3000
19	Q	209:	-104	24	-120	-88	-293	698	102	-525	121	-148	-252	122	-33	-356	-183	-96	-95	-167	-116	-260	-21	272	64	200	0	1200	-3000
20	G	198:	18	-77	-17	-130	-534	-113	-178	364	-60	-185	-231	-13	-224	-210	-299	14	-181	-274	-188	-232	-65	-90	-130	200	0	1200	-3000
21	Q	106:	-130	30	-78	-21	-593	122	21	-513	-42	-183	-219	59	-113	-99	-32	21	50	-334	-102	-204	-77	-34	25	200	0	1200	-3000
22	C	35:	-47	-253	-52	-170	538	-150	-172	-201	-188	-115	-66	-180	-31	-168	-183	-36	-18	-245	-179	35	-148	-162	-125	200	0	1200	-3000
23	G	21:	-77	-98	-51	-109	-172	-9	-66	-151	-51	-177	-161	-51	-72	-188	-140	-51	-61	-172	-82	-161	-93	-40	-114	200	0	1200	-3000
24	S	390:	-100	-341	53	-2	-82	90	25	-57	-153	-214	-33	[illegible]															000
25	C	326:	-76	-215	-78	-167	1043	-137	-156	-230	-130	-116	-7	[illegible]															000
26	W	326:	-258	-73	-133	-72	-238	37	-86	-147	-9	-78	-12	[illegible]															000

Aligning to Glycine at position 11 scores +6.5

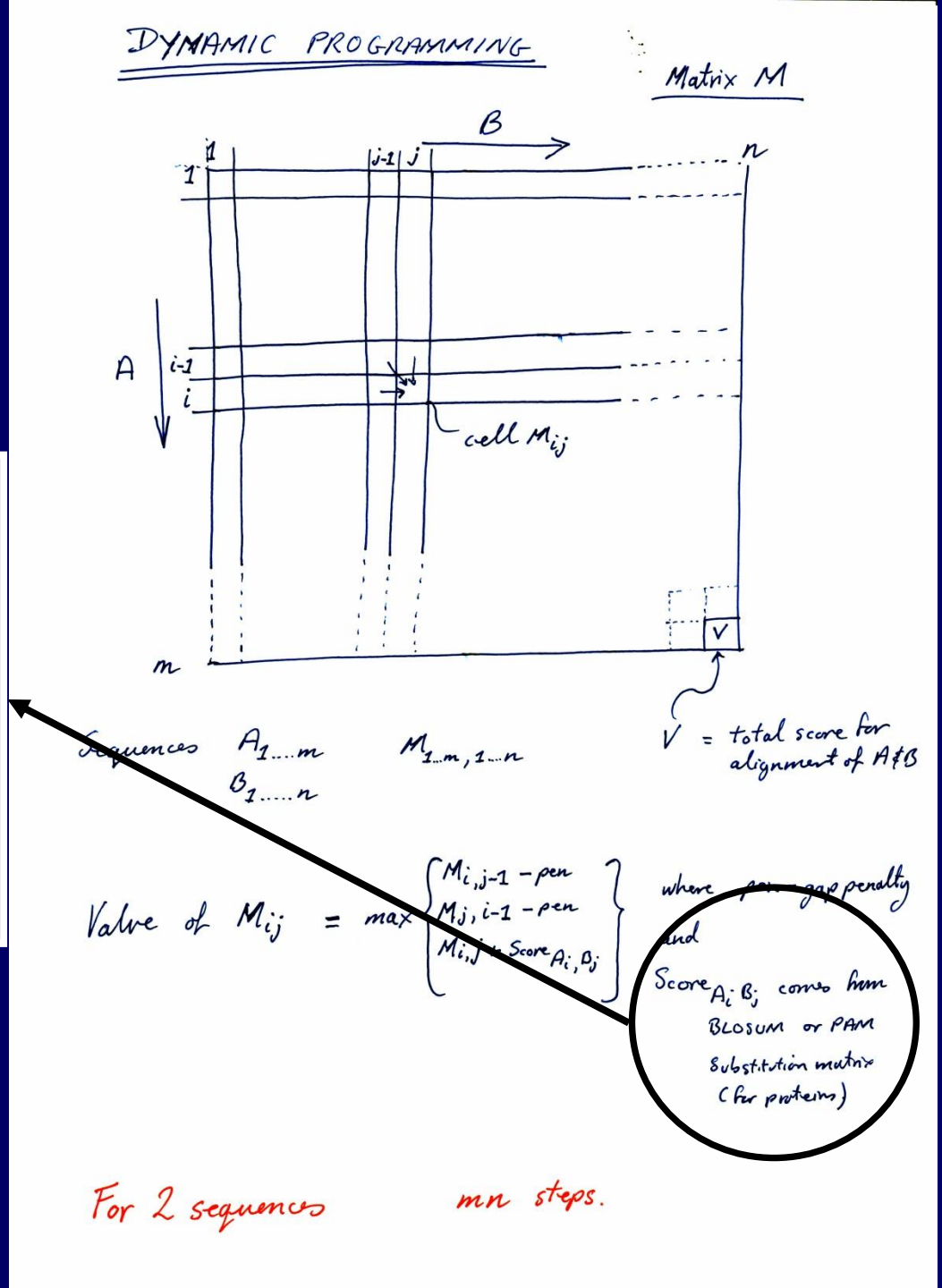
Aligning to Glycine at position 23 scores -1.51

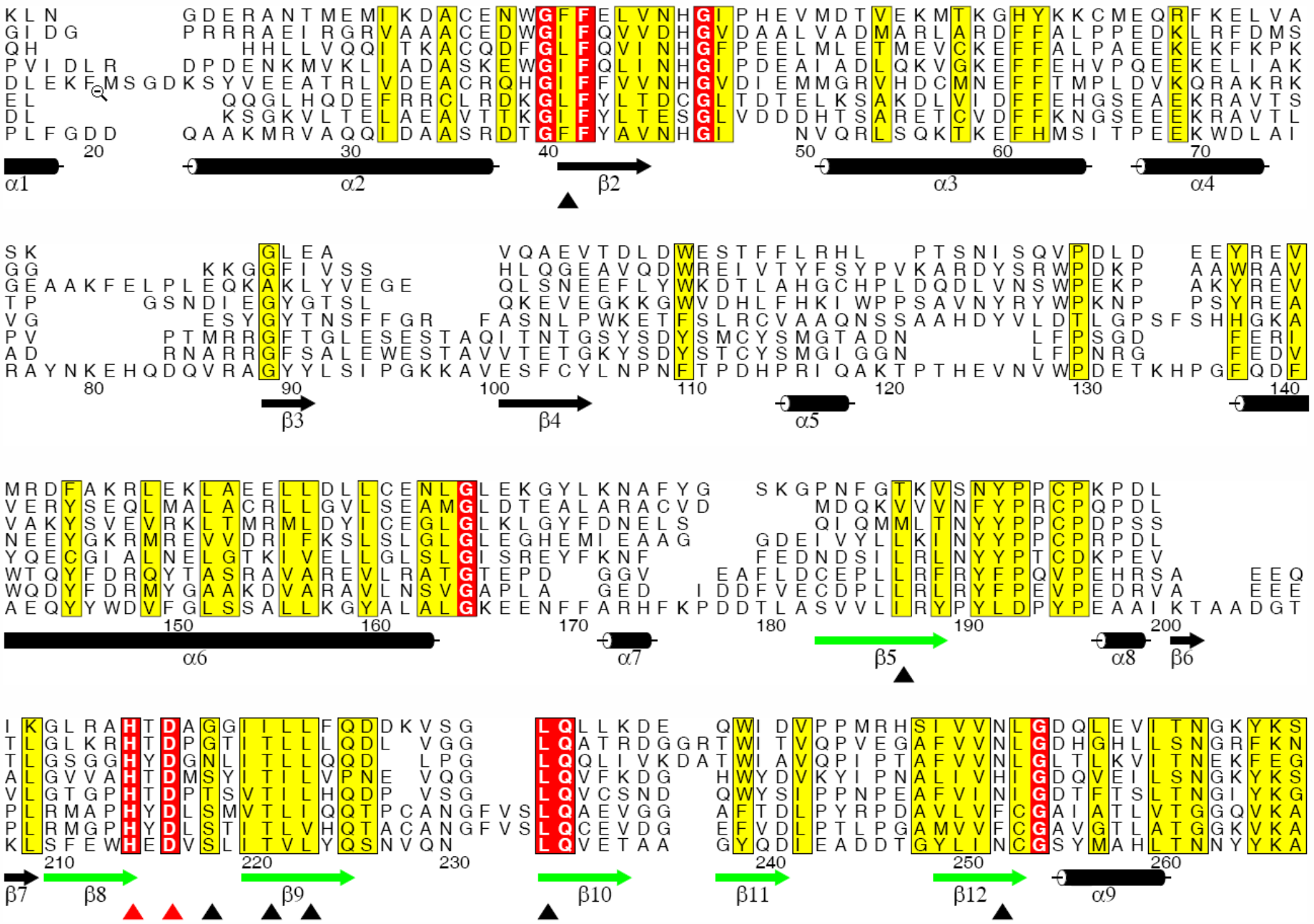
This emphasises position-specific features of the protein family

Compared to Gly-Gly score of 0.6 in the BLOSUM62 matrix.

When either A or B is a profile, the score comes from the profile rather than the BLOSUM matrix.

If A and B are both profiles, then the score is obtained by combining the scores from the two profiles (Exactly how is beyond this lecture)





ALSCRIPT output with colouring and secondary structure logos

(colour inkjet printer…) Roach *et al*, (1995)

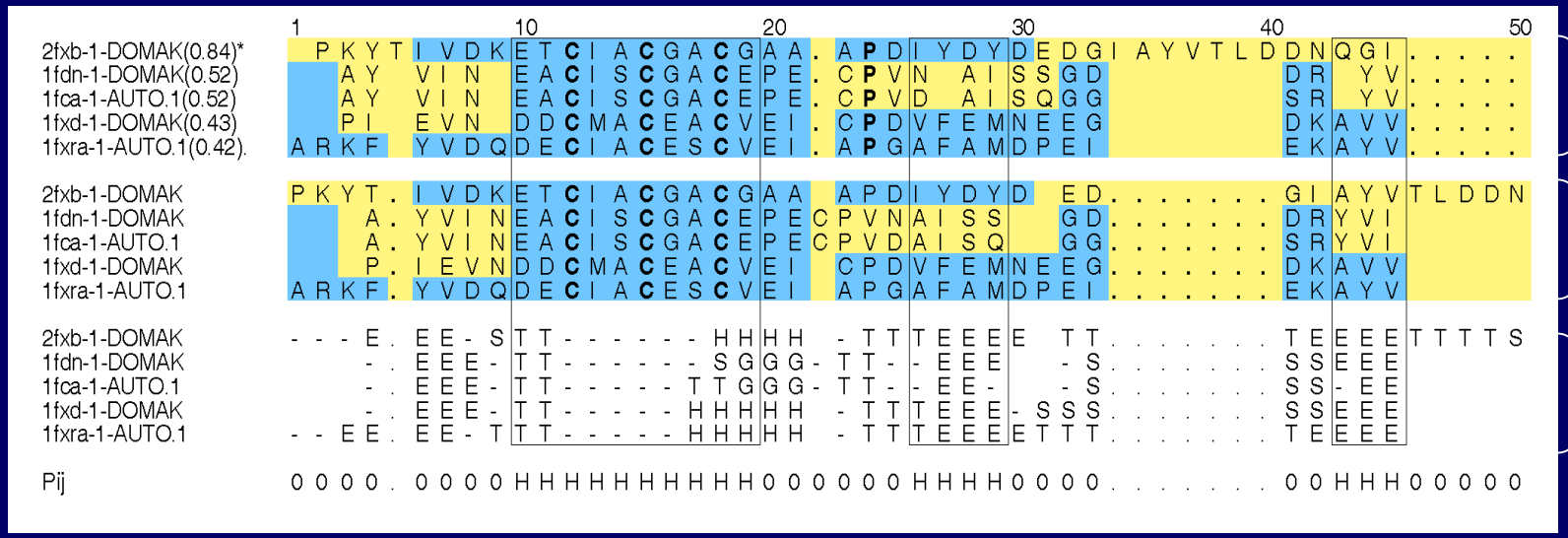
How good are alignments?

Use of reference alignments to see how well sequence alignments work

- OXBench – library of 672 multiple structure alignments
- Software to test how well different methods work

  - Raghava GP, Searle SM, Audley PC, Barber JD, Barton GJ. OXBench: a benchmark for evaluation of protein multiple sequence alignment accuracy. BMC Bioinformatics. 2003 Oct 10;4:47.

Comparison of Structural sequence alignment to sequence alignment



Sequence alignment

Reference structural alignment

Secondary structure for reference alignment

Boxed regions: STAMP reliably structurally aligned

BLUE highlighting: parts of the alignments that are the same.
YELLOW highlighting: parts of the alignment that is different.

Result of comparisons on alignments of 8 sequences or less

Grouped by percentage sequence identity (more on that later)

Hierarchical Methods

N-way DP with corner cutting →

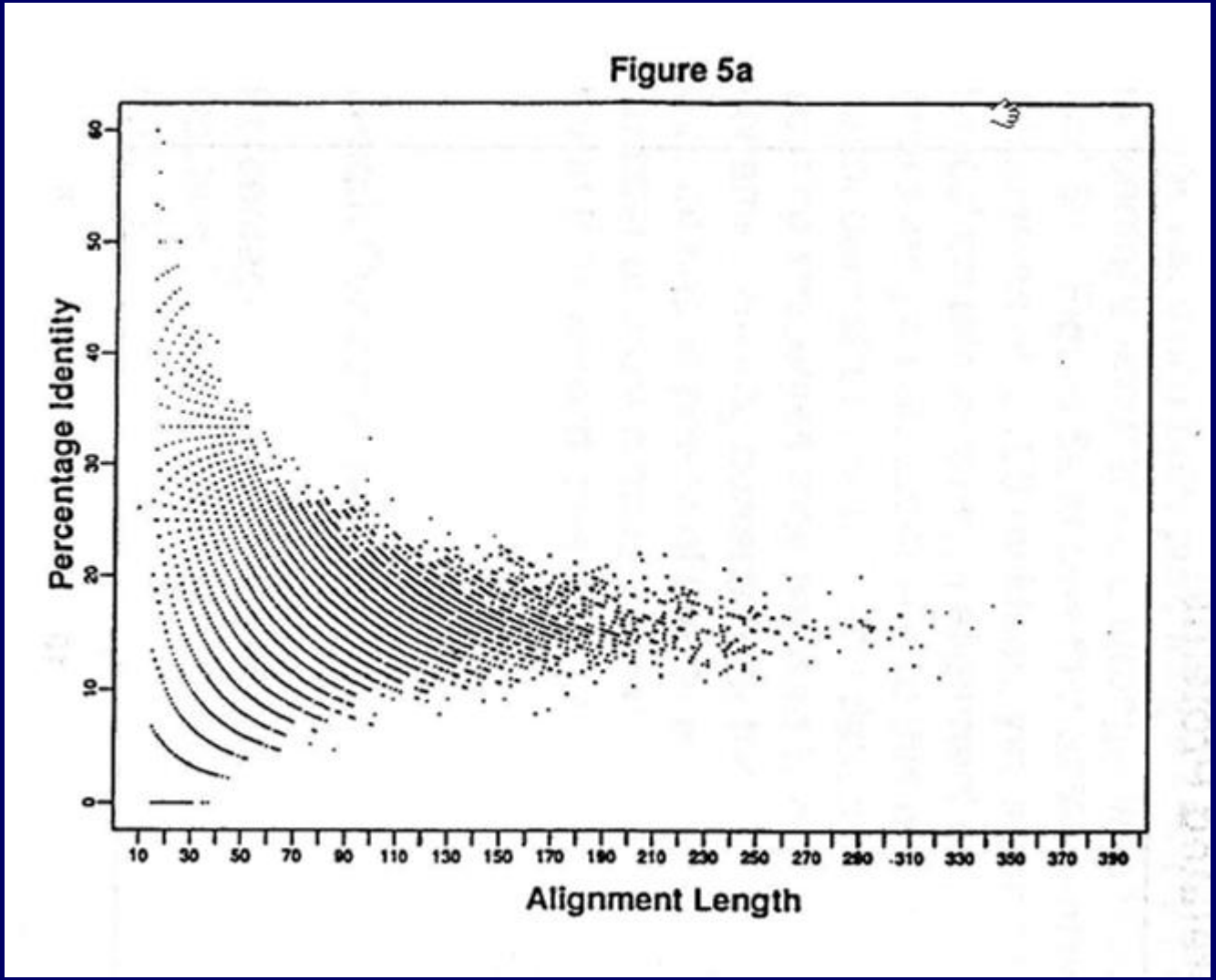
Methods	Percentage Identity Range					Overall
	0–10	10–20	20–30	30–50	50–100	
AMPS	22.2	62.2	81.5	91.3	99.0	89.68
CLUSTALW	21.4	57.0	79.3	91.2	99.0	88.94
PILEUP	25.9	59.5	78.4	90.2	99.0	89.00
PRRP	20.6	58.2	78.6	89.7	98.1	88.14
PIMA	17.4	56.6	78.7	90.1	99.0	88.46
DIALIGN	13.5	44.4	68.3	81.9	96.3	82.91
MSA	18.3	55.2	79.4	90.3	98.5	88.24
HMMER	6.1	13.2	27.8	55.9	89.4	66.20
T-COFFEE	23.1	69.0	87.2	93.3	99.2	91.39
N. Family	21	49	53	142	317	582

Table 10: The performance of methods on the MSA data set (families with $\leq$8 members.)

How similar do sequences need to be before we can align them reliably?

Percentage identity is strongly length dependent

Pair-wise sequence alignments of proteins known to be unrelated.



Barton, GJ, Proceedings of the CCP4 Study Weekend on Molecular Replacement (31 Jan-1 Feb, 1992)
There is a more recent ref with similar figure in it by Burkhard Rost, but I must find it!

Problems with percentage identity

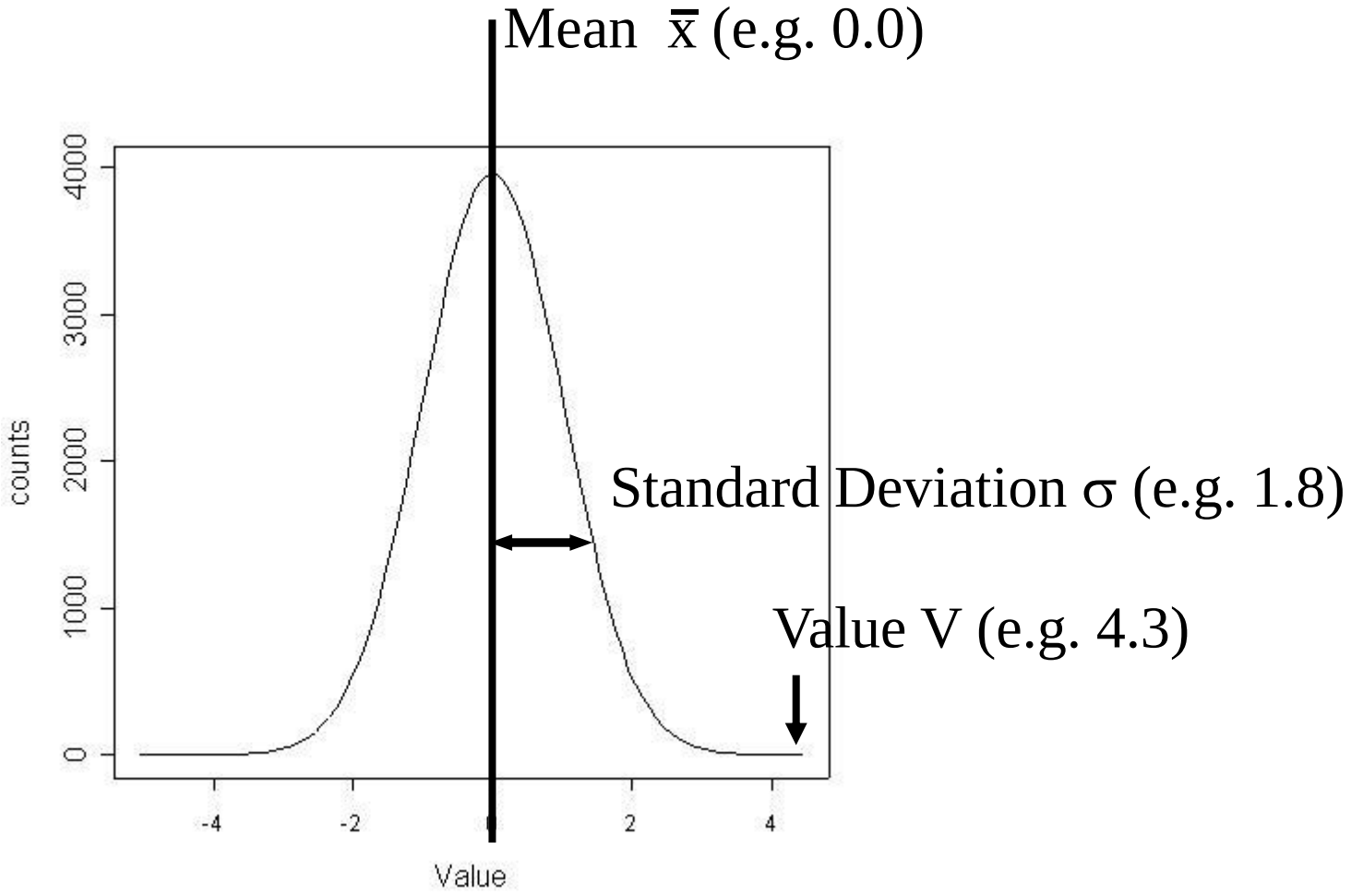
- Length-dependent
- Insensitive
- Dependent on the sequence alignment program and parameters
- Is a family of different scores…
  - Divide by length of shortest sequence
  - Divide by length of alignment
  - Divide by number of aligned positions etc.
    - See: Raghava, G.P.S. and Barton, G. J. Quantification of the variation in percentage identity for protein sequence alignments. BMC Bioinformatics. 2006 Sep 19;7:415.

Z-score compared to percentage identity

- Corrects for alignment length
- Is as sensitive as the alignment method
- Less sensitive to changes in the alignment method
- Only one way to calculate it

Z-score

- Align sequences and record score *S.*
- Shuffle order of amino acids in the sequences and re-align the pair. Record the score for this alignment, repeat 100 times.
- Calculate mean and Standard Deviation (sd) of shuffled sequence comparison scores.
- Z= (S-mean)/sd



Z-score = (Value – Mean)/(Standard Deviation)

= $(V - \bar{x}) / \sigma$

e.g. = (4.3-0.0)/1.8 = 2.39

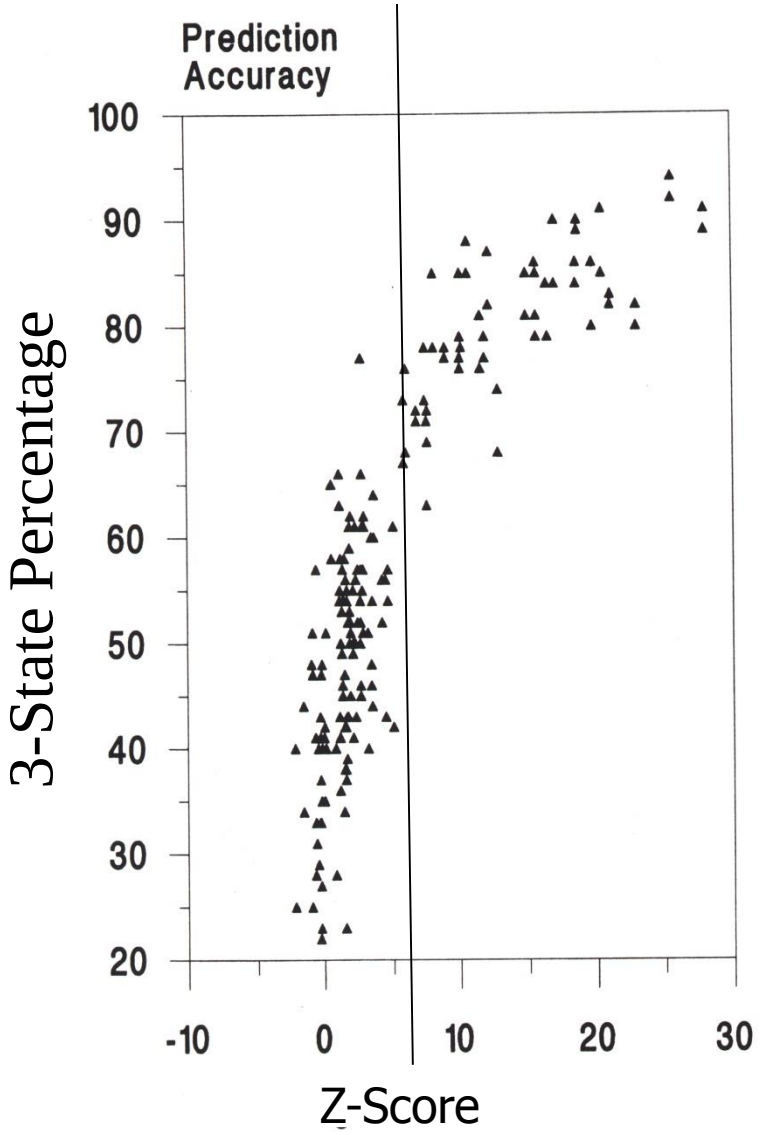


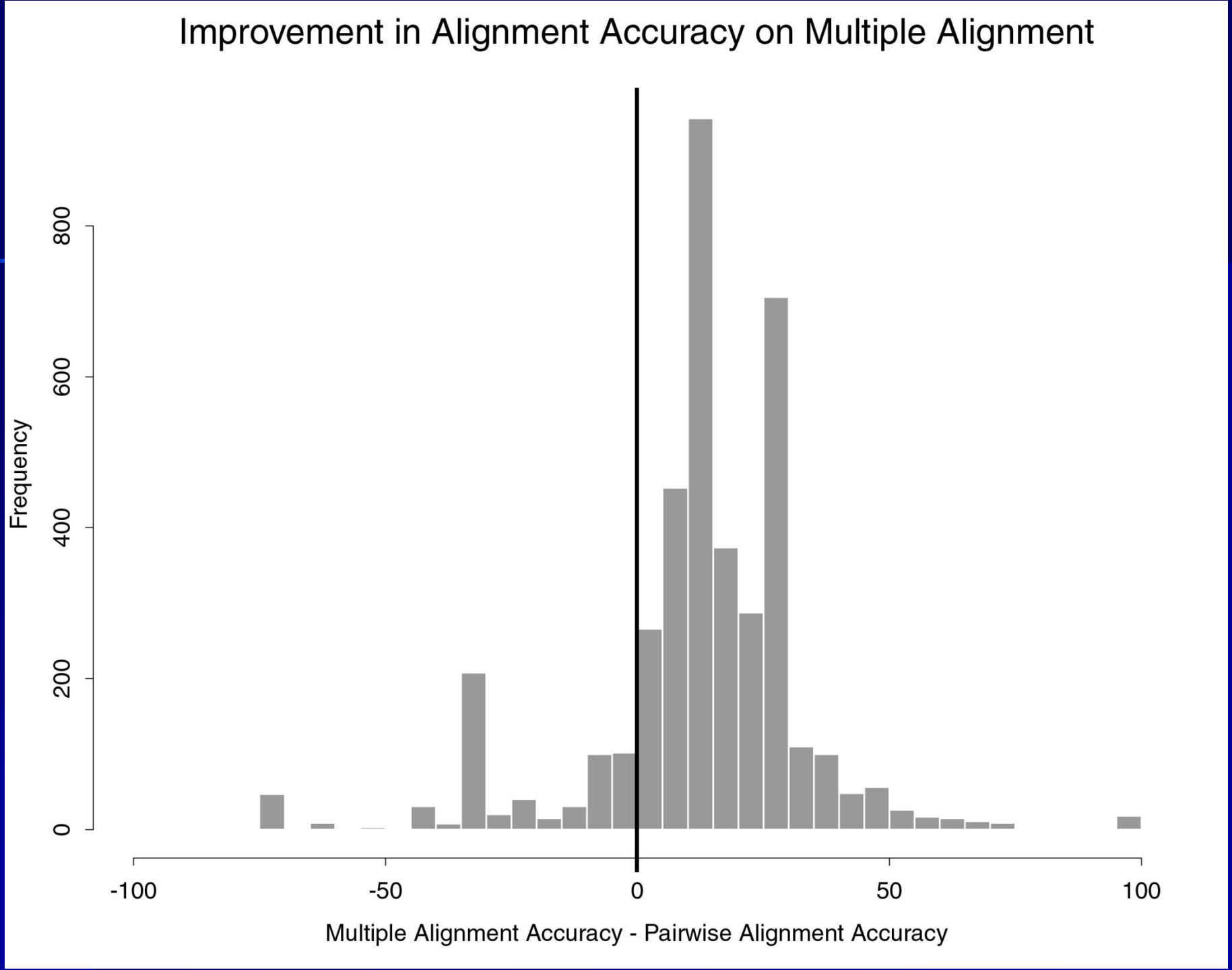
Fig. 2. The accuracy of secondary structure prediction by sequence alignment plotted against the alignment SD score to the homologous protein. One hundred and eighty-two predictions were made from pairwise alignment of the proteins in Table I.

Alignment accuracy judged by agreement of secondary structure

No poor alignments for similarity > 6 sigma.

Boscott, P. E., Barton, G. J. and Richards, W. G. (1993), *Prot. Eng.*,**6**, 261-266.

Alignment Accuracy Improves on Multiple Alignment

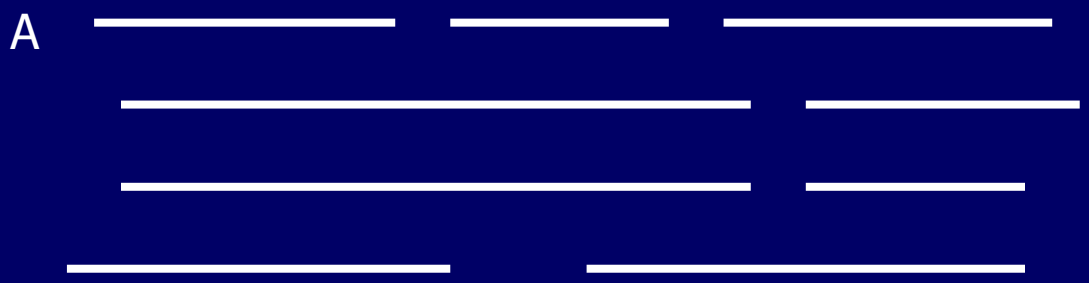

Improvement in Alignment Accuracy on Multiple Alignment
Frequency
0
200
400
600
800
-100
-50
0
50
100
Multiple Alignment Accuracy - Pairwise Alignment Accuracy

So, multiple alignments are on average more accurate than pair-wise alignments

Multiple alignments for different purposes

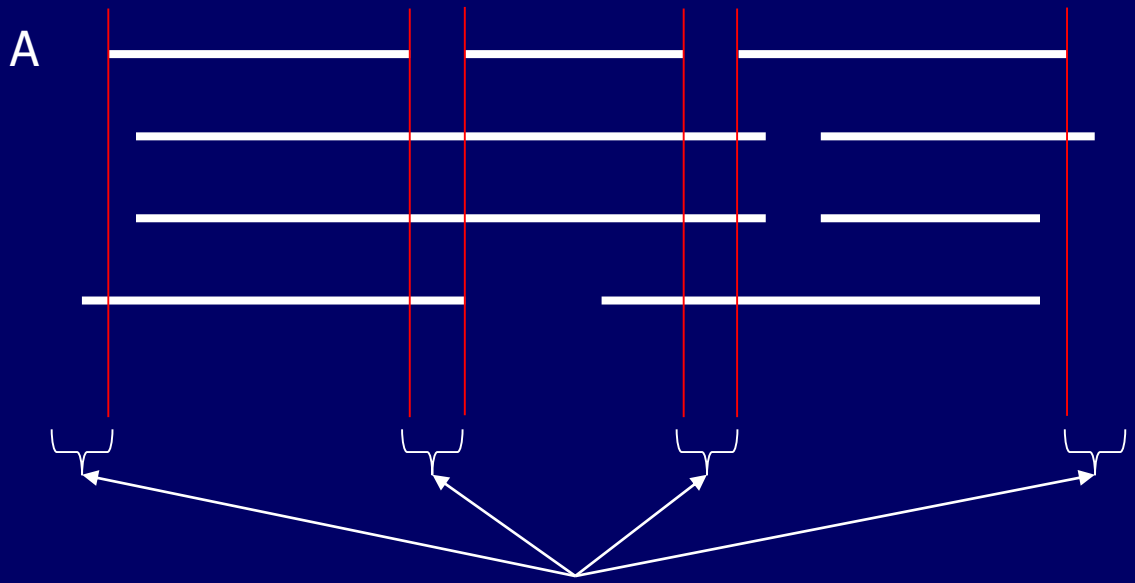
- Obtain best “full” alignment of N sequences.
  - Good starting point for most purposes.
- Obtain best alignment relative to one sequence
  - Best when subsequent analysis is focused on the first sequence.
  - Best approach for iterative profile searching since it prevents the alignment length growing longer than the sequence.

Multiple alignments for different purposes



Normal Hierarchical alignment: Gaps appear in the first sequence if needed

Alignment specific to sequence A.



Alignment relative to first sequence only: Regions of second and subsequent sequences aligned with gaps in first Sequence are sometimes deleted. e.g. JPRED output. and some PSIBLAST output.

Some uses of multiple Alignments

- Basis for sensitive profile searching of databases
- Identification of functional sites
- Phylogeny
- Presentation of sequence-related results
- Improved prediction of
  - Secondary structure
  - Disorder
  - Transmembrane regions
  - Almost any sequence-related property

Some uses of multiple Alignments

- Basis for sensitive profile searching of databases
- Identification of functional sites
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- Presentation of sequence-related results
- Improved prediction of
  - Secondary structure
  - Disorder
  - Transmembrane regions
  - Almost any sequence-related property

Why are multiple alignments useful for prediction?

- Evolution highlights amino acids important to maintaining the structure and function of a protein
- This information can be captured by visual analysis, or better, by machine learning techniques such as Artificial Neural Networks.
- This is what Day 3 of this course is about!

Jalview – a tool with which to tackle many of these analyses

Also good for RNA and DNA

Jim Procter

Suzanne Duce

Mungo Carstairs

Tochukwu (Charles) Ofoegbu

Jalview

www.jalview.org
twitter:@jalview

First developed in 1996

Funding: BBSRC and Wellcome Trust until 2019


Jalview